



Research Article

Trypanosomal infection rates in *Glossina pallidipes* in Bilbo village, Kamba District, Southern Ethiopia

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Abstract

This study was a cross-sectional study conducted at Bilbo village in Kamba district, Southwestern Ethiopia, from November 2018 to April 2019 with the aim of determining the infection rate of trypanosomes in *Glossina pallidipes*. A total of 384 *Glossina pallidipes* were captured using acetone and animal urine-baited non-gonococcal urethritis (NGU), bi-conical and sticky traps. Their organs (proboscis, salivary gland, and midgut) were dissected and microscopically examined. About 53.39% of *Glossina pallidipes* were captured through NGU, and the remaining 29.17% and 17.45% were captured via bi-conical and sticky traps, respectively. Out of the dissected specimens, 131 (34.1%) *Glossina pallidipes* were found positive for trypanosomes. Of this, 109 (28.38%) and 22 (5.73%) were female and male tsetse flies, respectively. The highest proportion of the tsetse fly (19.27%) was infected by *T. vivax*, followed by *T. congolense*, and *T. brucei*. There was a statistically significant difference in trypanosome infection rate among the sex ($P = 0.001$) and age ($P = 0.0024$) categories of *Glossina pallidipes*. The presence of *Glossina pallidipes* positive for trypanosomes might have contributed to bovine trypanosomosis in the study area. Hence, further studies should be undertaken in order to categorically prioritize the control of tsetse flies in the study area.

Keywords: *Glossina pallidipes*; *T. brucei*; *T. congolense*; *T. vivax*; Traps; Trypanosome infection rate

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1. Introduction

Tsetse flies are the cyclic vectors of trypanosomosis, a disease occurring mostly in rural areas and affecting agro-pastoral activities in rural communities (Feldmann et al., 2005). The fly has infested some 8–10 million km² of areas in 37 sub-Saharan African countries, including Ethiopia, corresponding to approximately one-third of Africa's total landmass (Cecchi & Mattioli, 2009). In Ethiopia, about 200,000 km² of areas in the western and southwestern parts of the

country are infested with tsetse flies. As a result, African Animal Trypanosomosis (AAT) remains a serious problem in the country (Alemu et al., 2007). Tsetse flies are confined to the southern, western, and southwestern regions of Ethiopia between longitudes 33° and 38° E and latitudes 5° and 12° N. Tsetse's infested areas lie in the lowlands and also in the river valleys of Baro, Akobo, Didessa, Abay (Blue Nile), Ghibe, and Omo (Langridge, 1976). The potential productive areas in the West and southwest parts of the country are infested by tsetse flies, mainly *Glossina morsitans*, *Glossina pallidipes*, *Glossina tachinoides*, and *Glossina fuscipes fuscipes* (Leta & Frehiwot, 2010; Denu et al., 2012; Desta et al., 2013). The most important trypanosome species affecting livestock in Ethiopia are *Trypanosoma congolense*, *Trypanosoma vivax*, and *Trypanosoma brucei* in cattle, sheep, and goats; *Trypanosoma evansi* in camels; and *Trypanosoma equiperdium* in horses (Getachew, 2005).

Glossina pallidipes is a moristan group of flies that is widely distributed in East Africa and is a major vector of animal trypanosomosis (Ouma et al., 2011). It is also present in other Eastern African countries such as Uganda and Somalia (Krafsur, 2009; Cecchi et al., 2014). The epidemiological importance of insect vectors increases with their age because they have a higher chance of becoming infected and more time to mature the infection (Leak, 1998).

The eco-distribution of the tsetse is determined by climate, the presence of vegetation, water, and the presence of blood meals (humans and animals). According to Rogers & Robinson (2004), Moore & Messina (2010), and Kleynhans & Terblanche (2011), the tsetse fly needs a habitat that is heavily impacted by ecological and climatic characteristics, including temperature, rainfall, soil and vegetation type, and other climate variables. Tsetse lives in habitats that provide shade for developing pupae and resting and breeding sites for adults (Rogers & Robinson, 2004). Their development is constrained by temperature and humidity, just like that of many invertebrates.

Temperature extremes, above 360 °C and below 100 °C lead to adult fly mortality because of starvation and water loss. Low humidity or moisture levels (directly related to precipitation) are also involved in fly mortality (Leak, 1998; Moore & Messina, 2010). Since there was no sufficient study on the trypanosome infection rate of *Glossina pallidipes* in this study area, this current preliminary study was aimed at obtaining baseline data on the trypanosome infection rate and associated factors influencing the occurrence of the infection of *Glossina pallidipes* in the study area.

2. Materials and Methods

2.1 Description of the study area

The study was conducted in Bilbo village, Kamba district, Gamo Zone, Southwestern Ethiopia. Kamba is bordered in the southwest by the South Omo Zone, in the West by Uba Derbretsehay, in the Northwest by Zala, in the Northeast by Deramalo, in the East by Bonke, and in the Southeast by the Dirashe Special Woreda.

The climatic conditions of the study area included a short rainy season (Late April to May), a long summer rainy season (from July to October), and a long dry season (late December to April). The study area reveals annual mean minimum and maximum temperatures of 11 to 160 °C and 22 to 31.20 °C, respectively, and an annual mean rainfall range of 723 to 1182mm. The study area reveals an elevation of 1021 meter above sea level. The study area encompassed long-grown grasses and small bushes. The dominant livestock species in the study area are cattle and goats. There are also some sheep, mules, donkeys, and chickens.

2.2 Study population

The study was conducted on *Glossina pallidipes*. A total of 384 *Glossina pallidipes* (266 female and 118 male tsetse flies) were captured using different types of traps. They were differentiated by sex and dissected for the determination of the trypanosome infection rate in the study area.

2.3 Study design

A cross-sectional study was conducted to assess the trypanosome infection rate of the *Glossina pallidipes* collected in the study area. The study was conducted from November 2018 to April 2019.

2.4 Sample size and sample size determination

The stratified sampling technique was used for the study of trypanosome infection rates in tsetse flies. Hence, a large number of *Glossina pallidipes* was captured, and at least 10% of the flies captured via traps were taken as study samples. Considering the tested population, which was over 5000, the sampling method was used with a 5% margin of error. The actual sample size has increased to 384 samples.

2.5 Materials and collection of tsetse flies

The study was conducted using non-gonococcal urethritis (NGU), bi-conical, and sticky traps deployed 100 meters apart on bushland. Hence, a total of 24 traps were deployed. All the traps were baited uniformly with octenol (1-oct-3-nel), acetone, and three-week-old cow urine (Dransfield et al., 1986). All odors were placed on the ground, about 30 cm upwind of the trap. The poles of the traps were greased to prevent predators, mainly ants. Traps were allowed to stay at the site of deployment for a period of 48 hours before collection. After 48 hours of deployment, the catchments of each trap were sorted by fly species and then counted, identified, and analyzed (Leak et al., 1998).

2.2.1 Sex determination

Tsetse flies were identified as male or female by examining the posterior end of the abdomen. The male fly has a lump on the ventral side of the abdomen (hypophgeum) at the posterior end but not in the female fly (FAO, 1992).

2.2.2 Age determination

In male tsetse, the age estimation was done according to the degree of wear or fraying observed on the hind margin of the wing. Flies were divided into one or more of the six categories outlined by Jackson (1946) and Challier (1965) based on the degree of wear. Female flies were age graded according to the contents of the uterus and the relative development of the four ovarioles. It was feasible to age the female tsetse flies by determining their ovarian age by performing tsetse dissection and evaluating the contents of the uterus as well as the relative size of the follicles in each of the two ovarioles and in each of the two ovules that constitute each ovary. According to Saunders (1962), each age category was divided into subgroups.

2.2.3 Determination of trypanosome infection rate

Wings were removed from the flies, and the degree of wing fraying was scored on a scale of 1 to 6 (Jackson, 1946). Then, freshly killed Tsetse flies were dissected under a dissecting microscope using 0.9% saline. Then, using the techniques of Lloyd & Johnson (1924), trypanosome infections in the tsetse flies were determined using a compound microscope at a magnification of 400. A cover slip was then placed on each area of the slide where the proboscis, salivary glands, or midgut were inserted.

Parasites found in the mid-gut, salivary glands, and mouth parts were regarded as Trypanozoon; “Trypanosoma brucei type), those located in the mouth parts and mid-guts were

Nanomonas (“Trypanosoma congolense type), and those found in the mouth parts only were put in the group of Duttonella (“Trypanosoma vivax type infection). The Infection rate (IR) was calculated using the following formula (Eq. 1):

$$\text{Infection rate} = \frac{\text{Number of Tsetse flies infected}}{\text{Total number of tsetse flies dissected}} \times 100 \quad (1)$$

2.6 Data analysis

The data was entered into a Microsoft Excel spread sheet to create a database and transferred to the Stata 9 software before analysis. The association between trypanosome infection rate and the assumed risk factors was tested with Pearson’s chi-square method. The trypanosome infection rate was calculated for all data using the number of infected tsetse flies divided by the total number of tsetse flies dissected over a given period of time and multiplied by 100.

3. Results and Discussion

3.1 Capturing performance of the traps

A total of 24 traps were deployed in the study area and 384 *Glossina pallidipes* were captured, of which 53.39% were caught by NGU traps, 29.17% by bi-conical traps, and 17.45% by sticky traps (Table 1). These findings indicate that the NGU trap was more effective than the other two trap types in the study area. The superior performance of the NGU trap aligns with the results of Dransfield et al. (1986), who also reported its higher efficiency in capturing *G. pallidipes* compared to bi-conical and sticky traps at Nguruman, Kenya. This consistency across different geographical settings suggests that the NGU trap may offer broader applicability in tsetse fly monitoring and control programs.

Table 1. *Glossina pallidipes* capturing capacity of deployed traps in the study area

Trap type	Total trap	<i>Glossina pallidipes</i> number	Proportion (%)
Nongonococcal urethritis	8	205	53.38
Bi-conical	8	112	29.17
Sticky	8	67	17.45
Total	24	384	100

3.2 Trypanosome infection rate in body tissues of *Glossina pallidipes*

The greatest proportion 74 (56.4%) of infection was detected in the proboscis of the fly, whereas 34 (26%) and 23 (17.6%) of trypanosomes were in the salivary gland and mid-gut of *Glossina pallidipes*, respectively (Table 2). With respect to trypanosome species, the infection rate associated with the *T. vivax* group (19.67%) was approximately twice as high as that of the *T. congolense* group (8.85%), while the *T. brucei* group showed the lowest prevalence (5.99%).

These findings are consistent with previous studies. Bitew et al. (2011) reported infection rates of 16.5% for *T. vivax*, 6.5% for *T. congolense*, and 0.5% for *T. brucei* in *G. pallidipes* from the Gojeb Valley, southeast Ethiopia. Similarly, Woolhouse et al. (1994) recorded infection rates of 6.2% for *T. vivax* and 3.1% for *T. congolense* in *G. pallidipes* from the Luangwa Valley, Zambia.

The predominance of *T. vivax* infections in the current study, particularly in the proboscis, may reflect the parasite's adaptation to mechanical transmission and its efficiency in exploiting this anatomical site. This suggests that *T. vivax* continues to be the dominant trypanosome species transmitted by *G. pallidipes*, with important implications for the epidemiology of animal trypanosomosis in the region.

Table 2. Trypanosome infection rate in body tissues of *Glossina pallidipes* captured at village of Kamba district

Body tissue	Total examined	Total positive	Infection rate (%)
Proboscis	158	74	56.4
Salivary gland	107	34	26.0
Mid-gut	119	23	17.6
Total	384	131	34.1

3.3 Trypanosome infection rate of *Glossina pallidipes*

The overall infection rate of *Glossina pallidipes* in Bilbo village, Kamba district, was 34.11% (Table 3). This accommodated *T. vivax*, *T. congolense*, and *T. brucie*. There was a significant difference between male and female tsetse flies ($\chi^2 = 18.139$; $P = 0.001$) in the proportion of infection with trypanosomes. Female tsetse flies revealed a higher trypanosomal infection rate (28.38%) than male tsetse flies (5.73%) in the study area. The trypanosomal infection rate was significantly higher in adult tsetse (30.73%) than young tsetse flies (3.38%) in the study area ($P = 0.0024$) (Table 4). Among the trypanosome species identified, *T. vivax* accounted for the highest infection rate (19.67%), followed by *T. congolense* (8.85%) and *T. brucei* (5.99%).

The higher infection rate in female flies compared to males is consistent with findings from Desta et al. (2013) in Amaro Special District, southern Ethiopia, where infection prevalence was 6.43% in females and 0.49% in males. This trend has been widely attributed to the longer lifespan of female flies, which increases the likelihood of acquiring infection during multiple blood meals (Mihok et al., 2008). The relatively low prevalence in males may be explained by the shorter average lifespan of trapped males, often less than 20 days. However, reports are not always consistent. For instance, Samdi et al. (2011) observed higher infection rates in males than females in Nigeria, while Zuk & McKean (1996) suggested that increased male involvement in sex-related

competition could elevate their exposure risk. These contrasting results highlight that host availability, fly longevity, and ecological variations may all contribute to infection dynamics across different geographical settings.

Table 3. The number of flies dissected and infection rate of *Glossina pallidipes* based on sex

Sex	TD	TP	Total flies infected by trypanosome species (%)			Infection rate
			<i>T. vivax</i>	<i>T. congolense</i>	<i>T. brucei</i>	
Female	266	109	64 (16.67)	28 (7.29)	17 (4.43)	109 (28.38)
Male	118	22	10 (2.60)	6 (1.56)	6 (1.56)	22 (5.73)
Total	384	131	74 (19.67)	34 (8.85)	23 (5.99)	131 (34.11)

TD=Total dissected, TP=Total positive

In the present study, *T. vivax* was the most prevalent trypanosome species detected. This finding is in line with previous reports from Ethiopia and other African countries, where *T. vivax* commonly dominates due to its ability to establish infections in the proboscis, facilitating efficient mechanical and cyclical transmission. The lower prevalence of *T. congolense* and *T. brucei* may be attributed to their more complex developmental requirements in the midgut and salivary glands, respectively. Collectively, these results underscore the epidemiological significance of *T. vivax* in the study area and reaffirm the need for targeted control strategies that consider both vector biology and parasite species distribution.

In the present study, adult tsetse flies exhibited a significantly higher infection rate (30.73%) compared to young flies (3.38%), which may be attributed to the longer lifespan and feeding frequency of adults, providing greater opportunities for trypanosome acquisition (Table 4).

Table 4. Trypanosome infection rate of *Glossina pallidipes* based on sex and age at Bilbo village

RF	Category	TE	TP	Trypanosome species (%)			IR	χ^2	P-value
				<i>T. vivax</i>	<i>T. congolense</i>	<i>T. brucei</i>			
Sex	Female	266	109	64 (16.67)	28 (7.29)	17 (4.43)	109 (28.38)	18.14	0.001
	Male	118	22	10 (2.60)	6 (1.56)	6 (1.56)	22 (5.73)		
Age	Young	70	13	6 (1.56)	4 (1.04)	3 (0.78)	13 (3.38)	9.20	0.0024
	Adult	314	118	68 (17.71)	30 (7.81)	20 (5.21)	118 (30.73)		
Total		384	131	74 (19.67)	34 (8.85)	23 (5.99)	131 (34.11)		

RF= Risk factors, TE= Total examined, TP=Total positive, IR= Infection rate

4. Conclusion

In conclusion, when compared to bi-conical and sticky traps, the NGU trap had the best ability to capture tsetse flies. As a result of the significant trypanosome infection rates found in *G. pallidipes*, this study also demonstrated the relevance of risk variables to the epidemiology of

African Animal Trypanosomiasis. Because trypanosome infection is the main impediment to livestock production in Bilbo village, Kamba district, southwest Ethiopia, the development of *G. pallidipes* positives for trypanosome species in the research area may be a factor. Therefore, additional entomological research should be conducted in order to prioritise tsetse fly management and address the severe trypanosomosis problem in the studied area.

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Conflict of Interest

There were, according to the authors, no conflicts of interest.

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